Amendments to the Specification:

Please replace the paragraph beginning on page 12, line 1, with the following rewritten paragraph:

~~As described above, B7L-1 can be used to separate cells expressing LDCAM, and,. In an alternative method, LDCAM or an extracellular domain or a fragment thereof can be conjugated to a detectable moiety such as ^{125}I to detect B7L-1-expressing cells. Radiolabeling with ^{125}I can be performed by any of several standard methodologies that yield a functional ^{125}I-LDCAM molecule labeled to high specific activity. Or an iodinated or biotinylated antibody against the B7L-1 region or the Fc region of the molecule could be used. Another detectable moiety such as an enzyme that can catalyze a colorimetric or fluorometric reaction, biotin or avidin may be used. Cells to be tested for B7L-1-expression can be contacted with labeled LDCAM. After incubation, unbound labeled LDCAM is removed and binding is measured using the detectable moiety.~~

As described above, B7L-1 can be used to separate cells expressing LDCAM. In an alternative method, LDCAM or an extracellular domain or a fragment thereof can be conjugated to a detectable moiety such as ^{125}I to detect B7L-1-expressing cells. Radiolabeling with ^{125}I can be performed by any of several standard methodologies that yield a functional ^{125}I-LDCAM molecule labeled to high specific activity. Or an iodinated or biotinylated antibody against the B7L-1 region or the Fc region of the molecule could be used. Another detectable moiety such as an enzyme that can catalyze a colorimetric or fluorometric reaction, biotin or avidin may be used. Cells to be tested for B7L-1-expression can be contacted with labeled LDCAM. After incubation, unbound labeled LDCAM is removed and binding is measured using the detectable moiety.

Please replace the paragraph beginning on page 13, line 1, with the following rewritten paragraph:

~~As described in Example 5, LDCAM is found on the PAE81BM cell line, which is an EBV transformed cell line. Thus, one example of such carrier use is to expose this cell line to a therapeutic agent/LDCAM conjugate to assess whether the agent exhibits cytotoxicity toward any EBV cancers.. Additionally, since LDCAM is expressed on dendritic cells and CD40L activated B cells that are important in antigen presentation, LDCAM is a useful carrier for targeting, identifying, and purifying these cells. Also, LDCAM/diagnostic agent conjugates may be employed to detect the presence of dendritic cells and B cells *in vitro* or *in vivo*. Example 6 demonstrates that human LDCAM mRNA, transcripts are found in human breast, retinal, fetal liver, spleen, fetal heart, lung, placenta, thyroid and lung carcinoma. Similar studies for expression of mouse LDCAM mRNA showed that mouse LDCAM mRNA is found in whole embryo, testes, lymphoid derived dendritic cells and triple negative cells. Since, LDCAM binds to itself, LDCAM can be used to study its functional role in these tissues.~~

As described in Example 5, LDCAM is found on the PAE81BM cell line, which is an EBV transformed cell line. Thus, one example of such carrier use is to expose this cell line to a therapeutic agent/LDCAM conjugate to assess whether the agent exhibits cytotoxicity toward any EBV cancers. Additionally, since LDCAM is expressed on dendritic cells and CD40L activated B cells that are important in antigen presentation, LDCAM is a useful carrier for targeting, identifying, and purifying these cells. Also, LDCAM/diagnostic agent conjugates may be employed to detect the presence of dendritic cells and B cells *in vitro* or *in vivo*. Example 6 demonstrates that human LDCAM mRNA, transcripts are found in human breast, retinal, fetal liver, spleen, fetal heart, lung, placenta, thyroid and lung carcinoma. Similar studies for expression of mouse LDCAM mRNA showed that mouse LDCAM mRNA is found in whole embryo, testes, lymphoid derived dendritic cells and triple negative cells. Since, LDCAM binds to itself, LDCAM can be used to study its functional role in these tissues.

Please replace the paragraph beginning on page 24, line 6, with the following rewritten paragraph:

~~To following describes expressing full length membrane-bound human LDCAM in CV1/EBNA cells. A vector construct for expressing human LDCAM was prepared by ligating the coding region of SEQ ID NO:1 into a pDC409 expression vector. The expression vector was then transfected in CV1/EBNA cells and LDCAM was expressed using techniques described in McMahan et al., *EMBO J. 10*:2821,1991.~~

The following describes expressing full length membrane-bound human LDCAM in CV1/EBNA cells. A vector construct for expressing human LDCAM was prepared by ligating the coding region of SEQ ID NO:1 into a pDC409 expression vector. The expression vector was then transfected in CV1/EBNA cells and LDCAM was expressed using techniques described in McMahan et al., *EMBO J. 10*:2821,1991.

Please replace the paragraph beginning on page 25, line 6, with the following paragraph:

~~Since the soluble human B7L-1 demonstrated binding to the murine lymphoma S49.1 (Example 2), a S49.1 expression library was screened for murine LDCAM cDNA clones. The process involved RT-PCR methodologies using the S49.1 cell line RNA and primers described in SEQ ID NO:7 and SEQ ID NO:8. These primers are based on a murine EST, discovered in a database and having homology to human LDCAM. The cDNAs were amplified by PCR using the primers, confirming the murine LDCAM is present in S49.1 cells.~~

Since the soluble human B7L-1 demonstrated binding to the murine lymphoma S49.1 (Example 2), a S49.1 expression library was screened for murine LDCAM cDNA clones. The process involved RT-PCR methodologies using the S49.1 cell line RNA and primers described in SEQ ID NO:5 and SEQ ID NO:6. These primers are based on a murine EST, discovered in a database and having homology to human LDCAM. The cDNAs were amplified by PCR using the primers, confirming the murine LDCAM is present in S49.1 cells.

Please replace the paragraph beginning on page 27, line 6, with the following paragraph:

~~Following detection of an appropriate antibody titer, positive animals are provided one last intravenous injection of B7L-1 in saline. Three to four days later, the animals are sacrificed, spleen cells harvested, and spleen cells are fused to a murine myeloma cell line, e.g., NS1 or preferably P3x63Ag8.653 (ATCC CRL 1580). Fusions generate hybridoma cells, which are plated in multiple microtiter plates in a HAT (hypoxanthine, aminopterin and thymidine) selective medium to inhibit proliferation of non-fused cells, myeloma hybrids, and spleen cell hybrids.~~

Following detection of an appropriate antibody titer, positive animals are provided one last intravenous injection of LDCAM in saline. Three to four days later, the animals are sacrificed, spleen cells harvested, and spleen cells are fused to a murine myeloma cell line, e.g., NS1 or preferably P3x63Ag8.653 (ATCC CRL 1580). Fusions generate hybridoma cells, which are plated in multiple microtiter plates in a HAT (hypoxanthine, aminopterin and thymidine) selective medium to inhibit proliferation of non-fused cells, myeloma hybrids, and spleen cell hybrids.

Please replace the paragraph beginning on page 27, line 13, with the following rewritten paragraph:

~~The hybridoma cells are screened by ELISA for reactivity against purified B7L-1 by adaptations of the techniques disclosed in Engvall et al., *Immunochem.* 8:871, 1971 and in U.S. Patent 4,703,004. A preferred screening technique is the antibody capture technique described in Beckmann et al., (*J. Immunol.* 144:4212, 1990) Positive hybridoma cells can be injected intraperitoneally into syngeneic BALB/c mice to produce ascites containing high concentrations of anti-B7L-1-L monoclonal antibodies. Alternatively, hybridoma cells can be grown *in vitro* in flasks or roller bottles by various techniques. Monoclonal antibodies produced in mouse ascites can be purified by ammonium sulfate precipitation, followed by gel exclusion chromatography. Alternatively, affinity chromatography based upon binding of antibody to protein A or protein G can also be used, as can affinity chromatography based upon binding to B7L-1.~~

The hybridoma cells are screened by ELISA for reactivity against purified LDCAM by adaptations of the techniques disclosed in Engvall et al., (*Immunochem.* 8:871, 1971) and in U.S. Patent 4,703,004. A preferred screening technique is the antibody capture technique described in Beckmann et al., (*J. Immunol.* 144:4212, 1990). Positive hybridoma cells can be injected intraperitoneally into syngeneic BALB/c mice to produce ascites containing high concentrations of anti-LDCAM monoclonal antibodies. Alternatively, hybridoma cells can be grown *in vitro* in flasks or roller bottles by various techniques. Monoclonal antibodies produced in mouse ascites can be purified by ammonium sulfate precipitation, followed by gel exclusion chromatography. Alternatively, affinity chromatography based upon binding of antibody to protein A or protein G can also be used, as can affinity chromatography based upon binding to B7L-1.